

1 **Thermophilic hydrolysis and acidification of activated sludge with a low organic carbon content under**
2 **different sludge concentrations**

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13 **Abstract**

14 The effect of residue activated sludge concentration on characteristics thermophilic hydrolysis and
15 acidification was investigated at the sludge concentrations of 10 g/L, 15 g/L and 20 g/L. Under steady state,
16 the volatile suspended solids (VSS) removal percentage in reactors fed with 10g/L, 15 g/L and 20 g/L residue
17 activated sludge was 10.8%, 10.1% and 10.7%, respectively. Soluble chemical oxygen demand (SCOD) was
18 highest in the reactor fed with 20 g/L sludge, while the amount of SCOD contributed by specific VSS reached
19 the peak when fed with 10 g/L sludge. Acidification rate was slightly decreased with increasing the sludge
20 concentration, which accounted for 74%, 72% and 69%, respectively and the ratio of various VFAs was
21 different. Ammonia and phosphate release was in proportion to the applied sludge concentration. At the
22 phylum level, *Firmicutes* was the most dominant phylum in three reactors. The relative abundance of
23 *Firmicutes* accounted for 19.6%, 21.2% and 23.5% fed with 10g/L, 15 g/L and 20 g/L sludge, respectively. At
24 the genus level, *Caloramator* and *Fervidobacterium* were dominant in three reactors, who might contribute
25 mostly to the hydrolysis and acidification process.

26

27 **Keywords:** Hydrolysis; Acidification; Sludge concentration; Microbial community; Thermophilic

28

29 **Introduction**

30 With the development of wastewater treatment, a large amount of residue activated sludge is produced and its
31 treatment/disposal usually costs nearly 25%-60% of operation cost of wastewater treatment plant [1].
32 Nowadays, landfilling, incineration and fermentation are main sludge treatment technologies. Especially,
33 anaerobic fermentation involves the degradation and stabilization of organic materials under anaerobic
34 conditions, and energy would be produced. In addition, hydrolysis and acidification products from anaerobic
35 digestion, especially soluble chemical oxygen demand (COD), can be used as organic carbon for enhancing
36 nitrogen and phosphorus removal from main stream wastewater treatment processes. Usually, hydrolysis and
37 acidification are limiting steps in anaerobic digestion of residue activated sludge [2]. Lots of efforts have been
38 endeavored to enhance sludge hydrolysis and acidification efficiencies or rates, such as alkaline treatment [3],
39 ultrasound treatment [4], and Fenton reagent reaction [5]. Physiochemical pretreatment processes costs too
40 much compared with biological augmentation, especially thermo-pretreatment processes.

41
42 Anaerobic digestion of sludge is usually carried out under mesophilic or thermophilic conditions.
43 Thermophilic anaerobic fermentation possesses high hydrolysis and acidification rate and efficiency, and also
44 can inactivate pathogens efficiently. Liu et al. showed that sludge hydrolysis was enhanced at high
45 temperatures [6]. For the dewatered sludge of 137400 mg/L total COD (TCOD), under mesophilic,
46 thermophilic and extreme thermophilic conditions, soluble COD (SCOD) concentrations were 25000, 25400
47 and 31000 mg/L, and volatile fatty acids (VFAs) concentrations were 2850, 6128 and 5463 mg/L, respectively.
48 Moreover, hydrolysis and acidification of sludge releases by-products of nitrogen and phosphorus, which can
49 be recovered as fertilizer if applicable. Therefore, enhancing thermophilic hydrolysis and acidification of
50 sludge is necessary to improve system performance through clarifying the possible underline mechanisms.
51 Sludge loading is an important factor affecting hydrolysis and acidification of sludge, which can be achieved
52 by adjusting sludge retention times (SRT) or sludge concentrations. Yuan et al. investigated the effect of SRT
53 and biomass concentration on non-fermentation of waste activated sludge, showing that VFA yields increased
54 with increasing SRTs and under the same SRT, VFA yields increased when the biomass concentration
55 decreased [7].

56
57 During thermophilic digestion, thermophiles, which play an important role in thermophilic hydrolysis and
58 acidification, may be influenced by operating conditions. Levén et al. found that the microbial community

59 structure at mesophilic and thermophilic temperatures was different. *Bacteroidetes* and *Chloroflexi* were
60 predominant at the mesophilic temperature while at the thermophilic temperature, *Thermotogae* was the major
61 group [8]. Yan et al. investigated the effect of heat-treatment for reduction of residue activated sludge on
62 microbial population and found that *Bacilli*, which included most of thermophiles, became the dominant class
63 in the community by the treatment [9]. Nevertheless, the effect of sludge concentration on microbial
64 community at thermophilic temperature is less understood.

65
66 Therefore, in this study, effect of sludge concentrations on hydrolysis and acidification under thermophilic
67 conditions was investigated, including volatile suspended solids (VSS) degradation, VFAs production and
68 release of protein, carbohydrate, nitrogen and phosphorus. In additions, effect of sludge concentration on
69 microbial community structure was also examined.

70

71 **Materials and Methods**

72 **Residue activated sludge**

73 The residue activated sludge was collected from the secondary settling tank of municipal wastewater treatment
74 plant in Kunming, China. The residue activated sludge was stored at 4°C after concentration. The main
75 characteristics of residue sludge were: pH of 6.8, SCOD of 83 mg/L, TCOD of 7400 mg/L, ammonia nitrogen
76 (NH₄-N) of 16.7 mg/L, ortho-phosphate (PO₄-P) of 0.51 mg/L, VSS of 17.5 g/L, and soluble protein and
77 carbohydrate of 236 mg/L and 5.2 mg/L, respectively.

78

79 **Anaerobic sequencing batch reactors (ASBRs)**

80 The effect of residue sludge concentrations on thermophilic hydrolysis and acidification was carried out in six
81 600 mL ASBRs with an effective volume of 500 mL. 10 g/L, 15g/L and 20 g/L residue activated sludge
82 concentrations were applied. Each sludge concentration contained two parallel reactors. All reactors were
83 incubated at 55°C and stirred at a rate of 150 r/min. 250 mL sludge was decanted every day and meanwhile
84 250 mL fresh sludge was dosed into each reactor to maintain both SRT and hydraulic retention time (HRT) of
85 2 days.

86

87 Nitrogen gas (N₂) was purged to remove the oxygen from the headspace of the reactor during dosing residue
88 sludge. 50 mL mix liquor was collected from the decanted samples. Before testing corresponding liquid

89 parameters, mix liquor was centrifuged at 8000 r/min for 5 min and then filtered with a 0.45 µm membrane
90 filter. Averaged results from the two parallel samples were presented.

91

92 **Analytical methods**

93 COD, NH₄-N, PO₄-P, suspended solids (SS) and VSS were measured according to standard methods [10] and
94 pH were measured by a pH meter (pH 30d, HACH, USA). Protein was measured according to [11].
95 Carbohydrate was measured by the sulfuric-phenol method [12].

96

97 VFAs (including acetic acid, propionic acid, iso-butyric acid, butyric acid, iso-valeric acid and valeric acid)
98 were measured by a gas chromatograph (GC-2014, Shimadzu, Japan) equipped with a flame ionization
99 detector and a capillary column. The carrier gas was N₂ at a flow rate of 50 mL/min, with a split ratio of 20 at
100 a flow rate of 1.1 mL/min in the column and a purge flow rate of 3.5 mL/min. The oven temperature was
101 increased proportionally from 80°C to 200°C within 10 min and the final holding duration was 2 min. The
102 temperatures of both injector and detector were 240°C. The samples was dosed with 1 µL formic acid to
103 adjusted pH to below 3.

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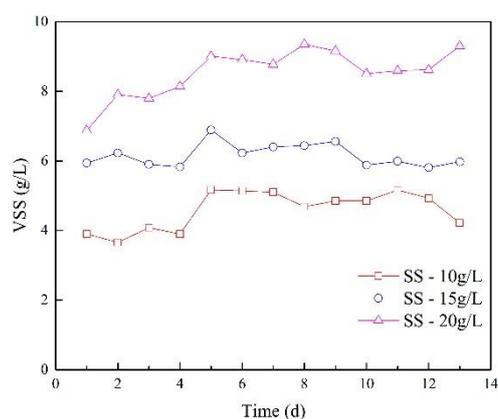
105 DNA was extracted from the anaerobic sludge in the six reactors using a Fast DNA Spin Kit (Laboratories,
106 Inc., Carlsbad, CA, USA) according to the manufacturer's instruction. After extraction, the DNA sample was
107 amplified by polymerase chain reaction (PCR). 16S rRNA high-throughput sequencing method was applied to
108 analyze microbial communities in all reactors.

109

110 **Results and discussion**

111 **Effect of sludge concentration on VSS hydrolysis**

112 Dynamics of VSS concentrations during the experiment are shown in **Fig. 1** After reaching steady state, the
113 VSS removal percentage was 10.8%, 10.1% and 10.7% with sludge concentrations of 10 g/L, 15 g/L and 20
114 g/L, respectively. Therefore, the VSS removal percentage was not affected by the residue sludge
115 concentrations. Banister et al. found that mass transfer efficiency could be enhanced at low sludge
116 concentrations, thus improving the VSS removal performance [13]. In our study, the applied sludge
117 concentration range did not affect system performance, and all condition might possess very good mixing
118 conditions.

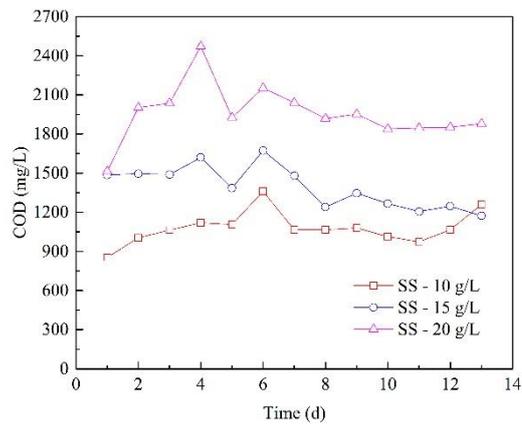


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120 **Fig. 1 Dynamics of VSS concentrations with different sludge concentrations**

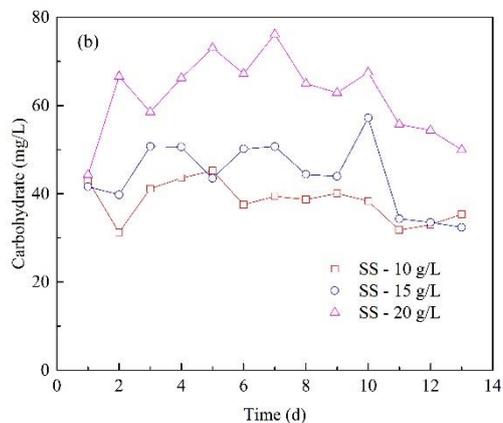
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122 Organic matters in residue activated sludge mainly exists in microbial cell and extracellular polymeric
 123 substances. These organic matters are hydrolyzed into small molecules substances, thereby increasing SCOD
 124 concentrations. Dynamics of SCOD corresponding to VSS degradation is shown in **Fig. 2** Under steady state,
 125 SCOD concentrations were 1053 mg/L, 1293 mg/L and 1903 mg/L at 10 g/L, 15 g/L and 20 g/L, respectively.
 126 SCOD increased with increasing residue activated sludge concentrations. Bouzas et al. found that with a SRT
 127 of 6 d at 20°C, a high concentration of residue activated sludge led to a high SCOD [14]. Xiong et al. also
 128 obtained that the highest SCOD was produced when feeding the highest VSS concentration in the range of
 129 4.57-23.78 g/L during hydrolysis of waste activated sludge [15]. Although increased sludge concentrations
 130 resulted in high SCOD concentrations, the specific yield coefficients of SCOD showed a different trend. The
 131 specific yield coefficients of SCOD were 262, 213 and 216 mg/g at 10 g/L, 15 g/L and 20 g/L, respectively.
 132 The highest specific yield of SCOD was obtained at 10 g/L, which was 23% and 21.3% higher than at 15 g/L
 133 and 20 g/L. Yuan et al. reported that longer SRTs resulted in higher solubilization of activated sludge and
 134 suggested that longer SRT enhanced the degradation of biomass [7]. According to the solubilization results
 135 obtained from this experiment, lower excess sludge concentration might also enhanced the degradation of
 136 biomass. The hydrolysis rates at 10 g/L, 15 g/L and 20 g/L conditions were 18.5%, 15% and 15.2%,
 137 respectively. These results were comparable with primary sludge fermentation results reported by Ucisik and
 138 Henze [16], where values of 15.9%-17.9% were obtained using an SRT of 6 d.



139
 140 **Fig. 2 Dynamics of SCOD with different sludge concentrations**

141
 142 Protein and carbohydrate accounted for more than 60% of the total organic matters in sludge [17]. During the
 143 hydrolysis, protein and carbohydrate were released. **Fig. 3** shows the effect of sludge concentrations on
 144 released soluble protein and carbohydrate concentrations. Under steady state, protein and carbohydrate in each
 145 reactor remained relatively stable, with protein concentrations of 119.9 mg/L, 142.9 mg/L and 185.3 mg/L,
 146 and carbohydrate concentrations of 36.6 mg/L, 52.4 mg/L and 67.4 mg/L in reactors fed with 10 g/L, 15 g/L
 147 and 20 g/L residue activated sludge, respectively. Chen et al. reported that protein and carbohydrate
 148 concentrations from 10.8 g/L waste activated sludge on the second day were 112.25 mg/L and 30.11 mg/L at
 149 pH 7.0 [2], which were similar to our results. Sludge concentration had almost the same effect on protein and
 150 carbohydrate concentrations as on SCOD. Both protein and carbohydrate were increased with increasing
 151 sludge concentrations. When the sludge concentration increased from 10 g/L to 20 g/L, concentrations of
 152 soluble protein and carbohydrate increased by 54.5% and 84.2%, respectively.



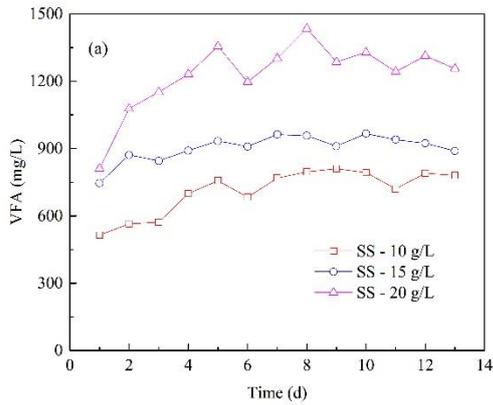
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 154 **Fig. 3 Soluble protein (a) and carbohydrate (b) concentrations with different sludge concentrations**
 155 **Effect of sludge concentrations on acidification processes**

156

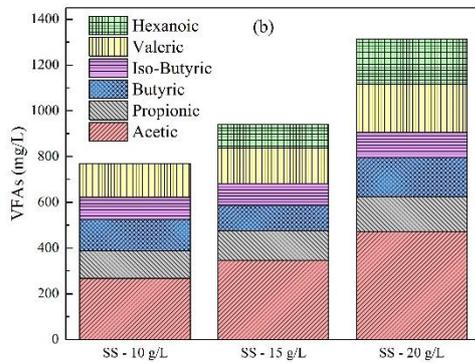
157 **Fig. 4a** shows dynamics of VFAs concentrations during the study period. The acidification rate was consistent
158 with sludge concentrations. The steady state VFAs concentrations were 780 mg/L, 935 mg/L and 1309 mg/L
159 in reactors fed with 10 g/L, 15 g/L and 20 g/L residue activated sludge, respectively. The specific VFAs yield
160 coefficients were 132 mg/g, 137 mg/g and 133 mg/g respectively, indicating that high sludge concentration
161 didn't inhibit acidification. Yuan et al. found that VFAs yield varied under different SRTs and the VFAs yield
162 increased with a decrease in sludge concentration for the sludge concentration range of 4.3-13 g/L [7]. For
163 example, at SRTs of 7 and 5 d, the VFAs yield increased by 19% and 16% when sludge concentration
164 decreased from 12.3 g/L to 4.8 g/L and from 12.4 g/L to 4.3 g/L, respectively. Skalsky et al. found that when
165 the solid concentration decreased from 2.6% to 0.43%, the VFAs yield increased by 50% [18]. In this study,
166 the acidification ratio (the proportion of VFAs to SCOD) was 74%, 72% and 69% at 10 g/L, 15 g/L and 20 g/L,
167 respectively. Therefore, only 5% acidification ratio increasing was obtained when the sludge concentration
168 decreased from 20 g/L to 10 g/L. These might be due to the high temperature applied in this study. Compared
169 to room temperature [18], thermophilic conditions might enhance mass transfer and improve the acidification
170 efficiency, thus reducing the inhibition of high sludge concentrations.

171

172 **Fig. 4b** shows components of VFAs in three reactors. Acetic acid, propionic acid, butyric acid, iso-butyric acid,
173 valeric acid and caproic acid were main VFAs generated by acidification. Acetic acid was dominant among all
174 VFAs, accounted for 35%, 41%, and 42% in reactor fed with 10 g/L, 15 g/L and 20 g/L sludge, respectively.
175 Liu et al. also reported that acetic acid was the dominant VFA under thermophilic conditions which followed
176 by propionic acid and butyric acid [6]. Propionic acid and butyric acid, on the other hand, accounted for 16%,
177 16% and 14%, and 18%, 13% and 15%, respectively. A low concentration of acetic acid was generated by the
178 reactor fed with 10 g/L sludge, while the maximum acetic acid was obtained by the reactor fed with 20 g/L.
179 Therefore, the produced VFA concentration could be affected by sludge concentration. Bouzas et al. found that
180 a high sludge concentration led to an increase of acetic acid, while propionic acid decreased [14]. Together, it
181 could be suggested that hydrolysis and acidification were both affected by residue activated sludge
182 concentration, and these two processes might be influenced by each other.



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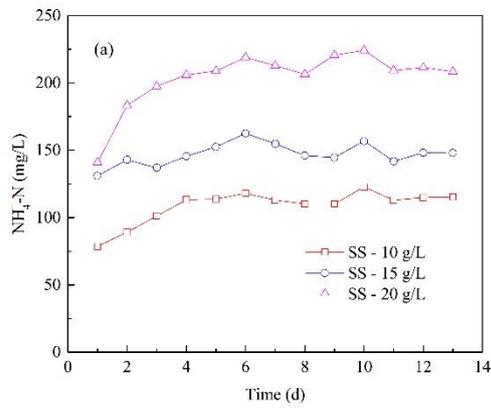
185 **Fig. 4 Dynamics of VFAs concentrations (a) and components of VFAs (b) with different sludge**
 186 **concentrations**

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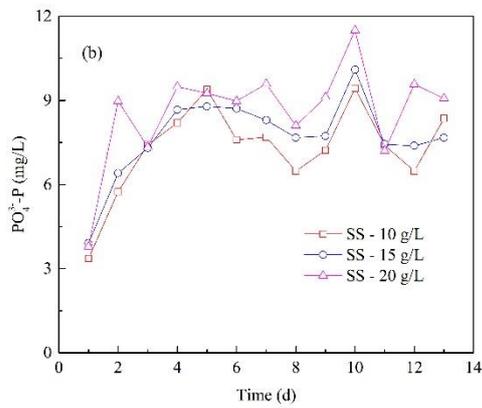
188 **Effect of sludge concentration on ammonia nitrogen and phosphate release**

189 **Fig. 5a** and **b** show the release of $\text{NH}_4\text{-N}$ and $\text{PO}_4\text{-P}$ under different sludge concentrations. $\text{NH}_4\text{-N}$ and $\text{PO}_4\text{-P}$
 190 both increased at the beginning. After three reaction cycles, concentration of $\text{NH}_4\text{-N}$ released was 114.2 mg/L,
 191 148.6 mg/L and 213.4 mg/L in reactors fed with 10 g/L, 15 g/L and 20 g/L sludge, respectively. Concentration
 192 of $\text{PO}_4\text{-P}$ was 5.8 mg/L, 8.3 mg/L and 10.6 mg/L, respectively. $\text{NH}_4\text{-N}$ and $\text{PO}_4\text{-P}$ concentrations were in
 193 proportion to residue activated sludge concentrations. By increasing residue activated sludge concentrations
 194 from 10 g/L to 15 g/L and then to 20 g/L, concentration of $\text{NH}_4\text{-N}$ increased by 30.1% and 86.9%. The
 195 corresponding $\text{PO}_4\text{-P}$ concentration increased was 43.1% and 82.8%, respectively. Pitman et al. found that the
 196 $\text{PO}_4\text{-P}$ release potential was consistent with SS concentrations [19]. Moreover, concentrations of $\text{NH}_4\text{-N}$ and
 197 VFAs (**Fig. 5c**) had a good linear relationship ($R^2=0.94$), which might be used to reflect the acidification

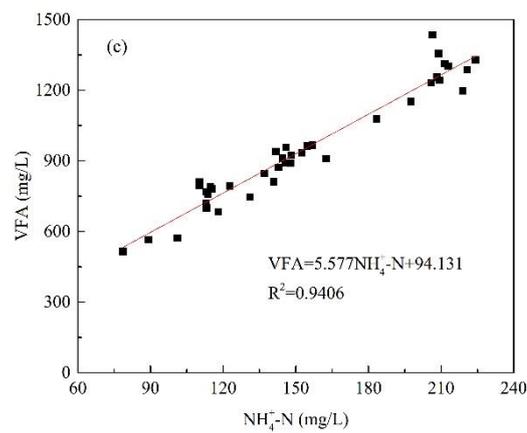
198 process. This might be due to that protein was the dominating organic carbon in sludge, which released
199 nitrogen during acidification.



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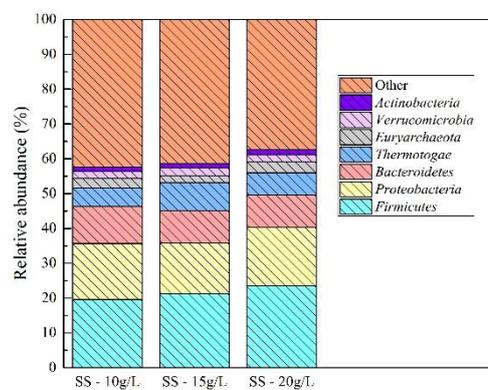
203 **Fig. 5 The release of $\text{NH}_4\text{-N}$ (a) and $\text{PO}_4\text{-P}$ (b) with different sludge concentrations and relationship**

204 **between $\text{NH}_4\text{-N}$ and VFA (c)**

205

206 **Microbial community for systems with different sludge concentrations**

207 Microbial community structure of three reactors fed with different sludge concentration was identified (**Fig. 6**).
 208 At the phylum level, *Firmicutes* was the dominant phylum, accounting to 19.6%, 21.2% and 23.5% in reactors
 209 fed with 10 g/L, 15 g/L and 20 g/L sludge, respectively. *Proteobacteria* was the followed dominant phylum,
 210 accounting to 16.0%, 14.6% and 16.8%, respectively. *Bacteroidetes*, the third dominant phylum, accounted to
 211 10.6%, 9.2% and 9.3%, respectively. At the genus level, *Caloramator* (belonged to *Firmicutes* phylum) and
 212 *Fervidobacterium* were dominant in three reactors. The relative abundance of *Caloramator* increased with
 213 increasing sludge concentrations, accounting to 9.8%, 11.0% and 13.1% when fed with 10g/L, 15 g/L and 20
 214 g/L residue activated sludge, respectively.



215
 216 **Fig. 6 Microbial community structure at phylum level with different sludge concentrations**

217
 218 Microorganisms within *Firmicutes*, *Proteobacteria* and *Bacteroidetes* were widely distributed throughout not
 219 only mesophilic conditions but also thermophilic conditions [8, 20]. Digestion at thermophilic temperature
 220 provides higher efficiency in the degradation of organic matters [21, 22]. Yan et al. reported that thermophilic
 221 proteases which contribute to the hydrolysis of residue activated sludge could be secreted by thermophiles and
 222 the overall protease activities measured at thermophilic temperature was significantly higher than that at 28 °C
 223 [9]. Tan et al. reported that *Firmicutes* and *Proteobacteria* were found when sludge was treated at 60 °C [20],
 224 which indicated that these bacteria might be heat-resistant. *Firmicutes* have the ability to metabolize substrates
 225 such as proteins, lipids, celluloses, sugars and amino acids by producing extracellular enzymes [8]. *Firmicutes*
 226 was the typical phylum in all reactors, indicating that it was probably responsible for organic carbon and
 227 proteins reduction. What's more, the relative abundance of *Firmicutes* increased with increasing sludge
 228 concentrations, suggesting that it could adapt well to the high organic loading applied in this study.
 229 *Proteobacteria* was typical phylum in hydrolysis and acidification of ultrasonic-pretreated wasted activated

230 sludge [23]. *Bacteroidetes* could convert proteins and carbohydrates to propionic acid and acetic acid as
231 primary products during anaerobic digestion of sludge [24]. Therefore, *Bacteroidetes* might be also involved
232 in the fermentation process of soluble proteins and carbohydrates.

233

234 **Conclusions**

235 Effect of sludge concentrations on thermophilic hydrolysis and acidification of sludge was investigated.
236 Sludge concentration had little effect on VSS removal percentage. Soluble COD, protein and carbohydrate
237 concentrations increased with increasing the sludge concentration. The acidification was consistent with the
238 sludge concentration and the highest VFAs yield was obtained in reactor fed 20 g/L residue activated sludge.
239 Thermophiles were found in all reactors and at the phylum level, *Firmicutes* was the most dominant phylum in
240 three reactors, whose relative abundance increased with increasing sludge concentrations.

241

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