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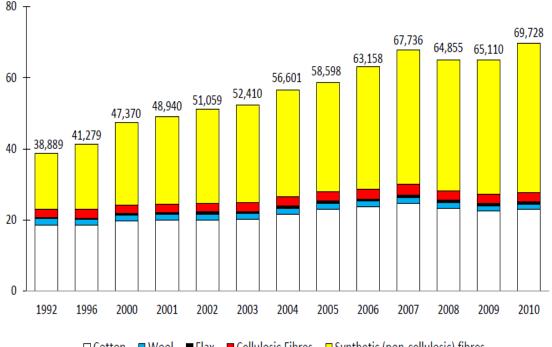
# Cellulase Production via Solid State Fermentation on Textile Waste

HU Yunzi

5th International Conference on Sustainable Solid Waste Management Athens, 2017

### **Project background-Textile consumption**

- In the decade of 2001-2010, the textile consumption expanded by 42 43%
- In 2011-2014, annual textile consumption reached **80 90 million tonnes**



#### **Textile wastes: 10-20% of all textile products**

(ITP/109/15TP)

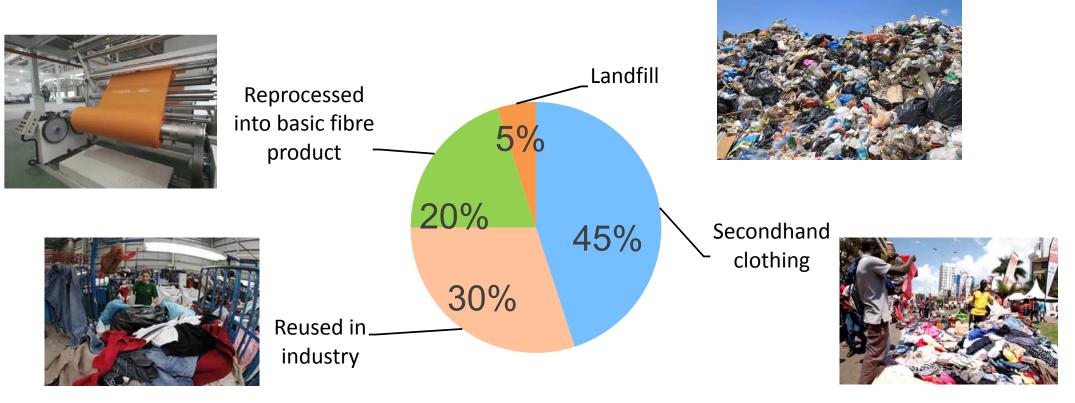
Generation wast (million tonn	e	Textile waste per capita (kg/year)
China	26.0	19.2
UK	1.0	15.6
US	12.4	32.0
HK	0.17	23.7

□ Cotton ■ Wool ■ Flax ■ Cellulosic Fibres □ Synthetic (non-cellulosic) fibres

Source: China Association of Resource Comprehensive Utilization; Waste & Resources Action Programme (UK); SMART textile recycling (US); Department EP of Hong Kong

# Where did they go?

- Landfill: 85% of the total textile solid waste
- Recyclable percentage: 95% of landfilled textile waste
- Currently recycling percentage: **14-15%**

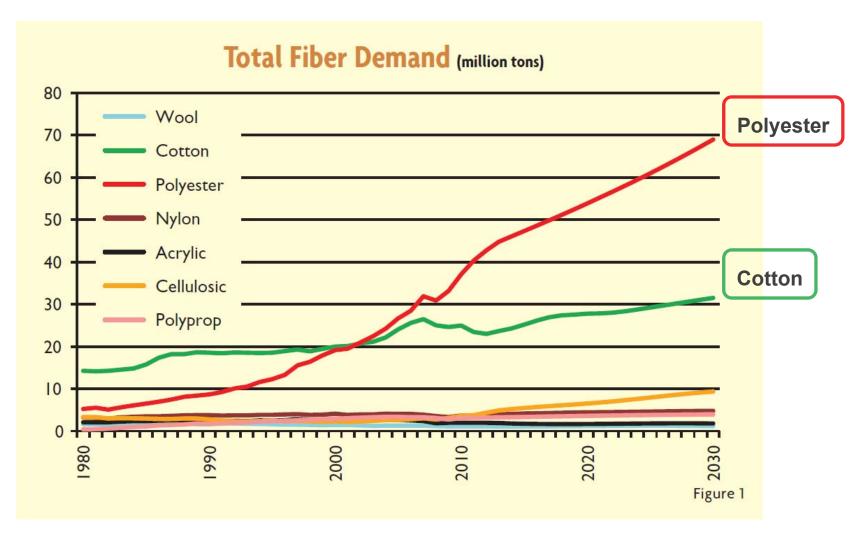


Source: U.S. Environmental Protection Agency and SMART Association, British Material Recycling Association

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# **Project background-Textile consumption**

• Cotton and Polyester: Top 2 in Fibre Demand

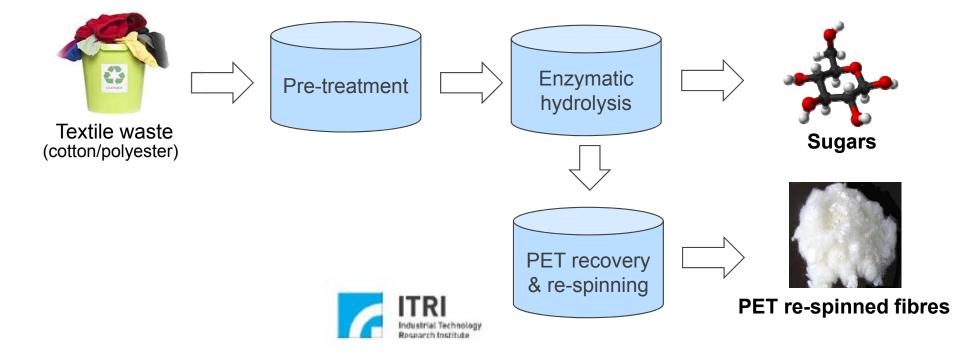


Source: Textile World, Man-Made Fibers Continue To Grow

# **Project introduction**

### We propose a sustainable textile waste recycling strategy in HK:

- **1. A novel approach of textile waste treatment via biological method**
- 2. Recovery of glucose from textile wastes through enzymatic hydrolysis
- 3. Separate the **PET** fiber from textile waste and reuse it in textile industry

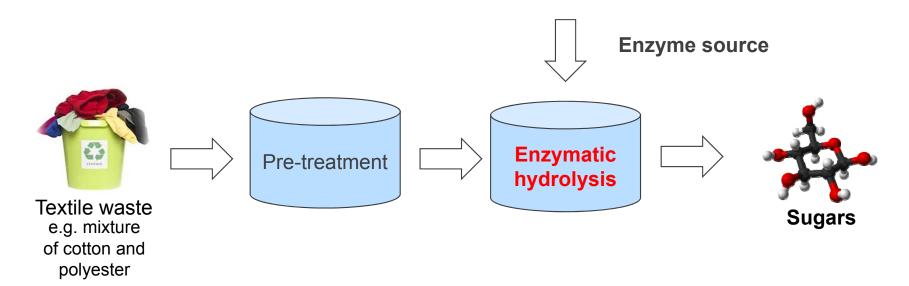


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### **Research target**

### Fungal cellulase production on textile waste

(Conducted by Dr. Carol Lin from CityU and Dr. Du Chenyu from the University of Huddersfield)



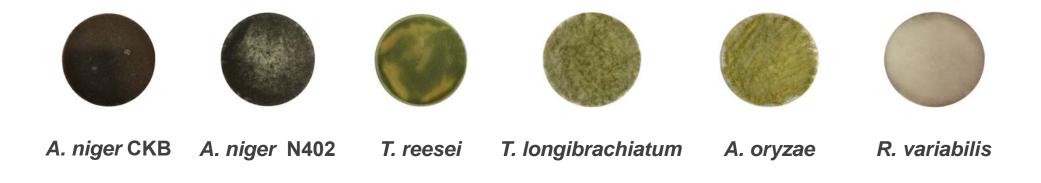
Materials	Dye	Pre-treatment	Source of enzymes
<ul> <li>100% Cotton</li> <li>100% Polyester</li> <li>Cotton/PET blend (80/20, 60/40, 40/60)</li> <li>Jean</li> </ul>	<ul><li>Reactive</li><li>Disperse</li><li>Indigo</li></ul>	<ul><li> Alkaline</li><li> Milling</li><li> Autoclave</li></ul>	<ul> <li>Fungal enzymes (solid state fermentation: SSF)</li> </ul>

Substrate: different types of textile fabrics (from H&M)

	A REAL					
Component	Cotton 100%	Cotton 80% PET 20%	Cotton 60% PET 40%	Cotton 40% PET 60%	PET 100%	Jeans (Cotton 99%, Elastane 1%)
Dye	Reactive dyestuff	Reactive dyestuff	Reactive dyestuff	Reactive dyestuff	Disperse dyestuff	Indigo dye

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#### **Cellulase producing fungal strains**



Aspergillus niger CKB: Isolated from natural environment (provided by Dr. Diannan Lu, Tsinghua University)

Aspergillus niger N402: From Prof. David Archer in the University of Nottingham in the United Kingdom

T. reesei: Trichoderma reesei ATCC 24449

*T. longibrachiatum: Trichoderma longibrachiatum (*Prof. Colin Webb, The University of Manchester, United Kingdom)

*R. variabilis: Rhizomucor variabilis* (provided by Tsinghua University)

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Substrate: 100% cotton fabric

Day 7

A. niger CKB

Moisture: 65-85%

**Duration: 7 days** 

Day 0

**Before SSF** 

**Temperature: 28°C** 

Supplemented nutrient: yeast extract 2.5% (w/w)

### **Cellulase production via SSF on textile waste**

### **1. Screen of fungal strains**

Cutting

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A. oryzae

A. niger N402

R. variabilis



T. longibrachiatum



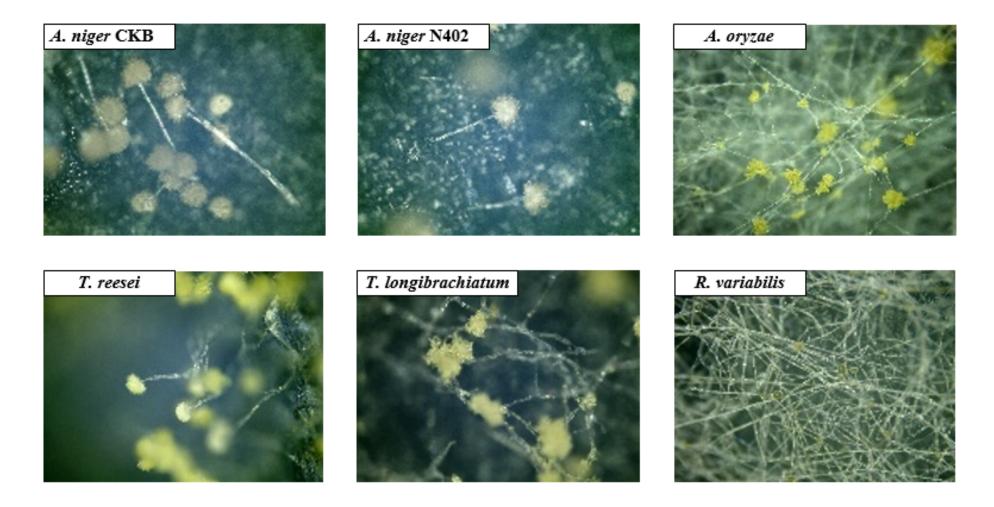
T. reesei

(ITP/109/15TP)

 $0.8 \times 0.8 \text{ cm}^2$ 

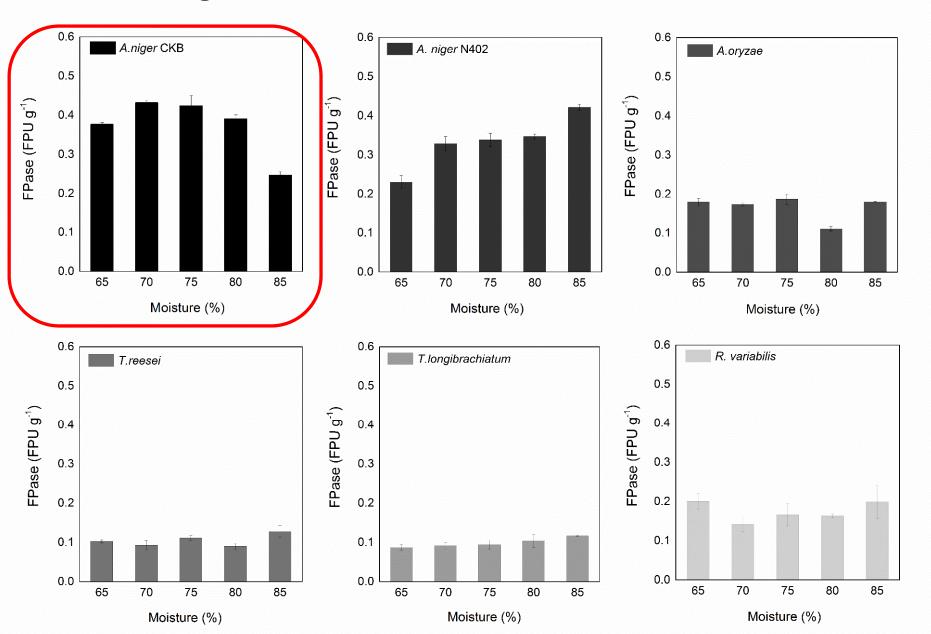
#### **1. Screen of fungal strains**

#### **Microscope detection**



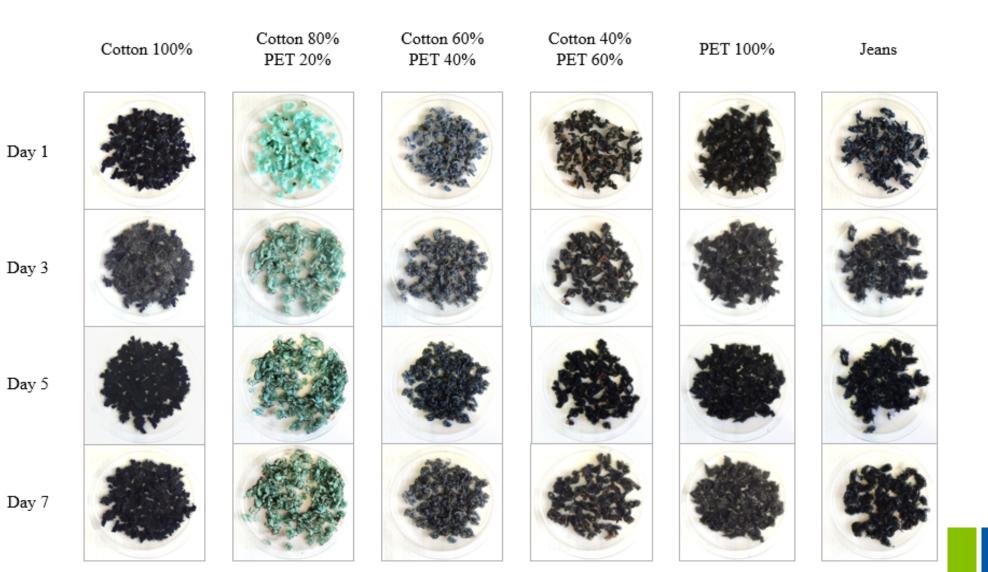
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### 1. Screen of fungal strains



#### 2. Comparison of different textile fabrics

Substrate: 6 types of textile fabrics; Moisture: 75%; Temperature: 28°C; Yeast extract: 2.5 w/w%

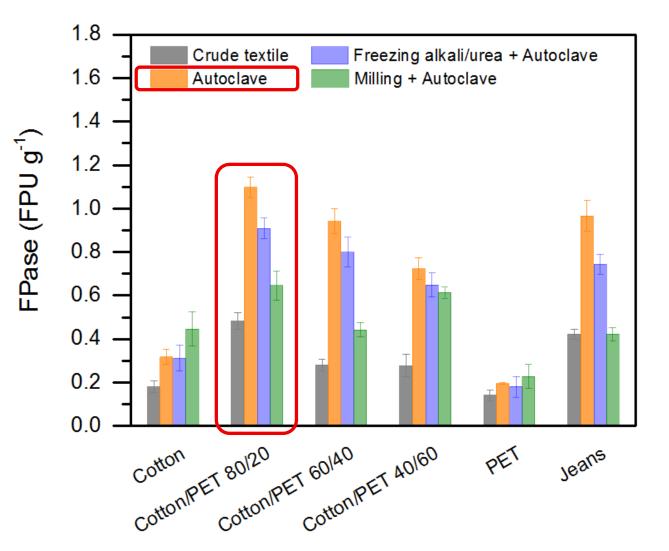


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#### 2. Comparison of different textile fabrics

Pretreatments

- 1. Autoclave (121°C for 15 min)
- 2. Freezing alkali/urea (Soaking at -20 °C for 6 h)
- 3. Milling (powder form, < 1 mm<sup>2</sup>)



Polytechnic University 香港理工大學

Pretreatments were conducted by our collaborators Dr. Shao-Yuan Leu and Dr. Hao Liu in Hong Kong Polytechnic University

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#### 4. Optimization of fermentation condition by Response Surface Methodology



(ITP/109/15TP)

Response: Cellulase activity (FPU g<sup>-1</sup>)

Numeric factor	Unit	Low value	High value	-alpha	+alpha
A. pH	-	5	7	4	8
B. Yeast extract	w/w %	1	4	0	5.5
C. Inoculum size	Spores g <sup>-1</sup> textile	1.6E+007	4.6E+007	1E+006	6.1E+007
D. Moisture	%	60	80	50	90

#### ANOVA for Response Surface Quadratic Model

#### Analysis of variance table [Partial sum of squares - Type III]

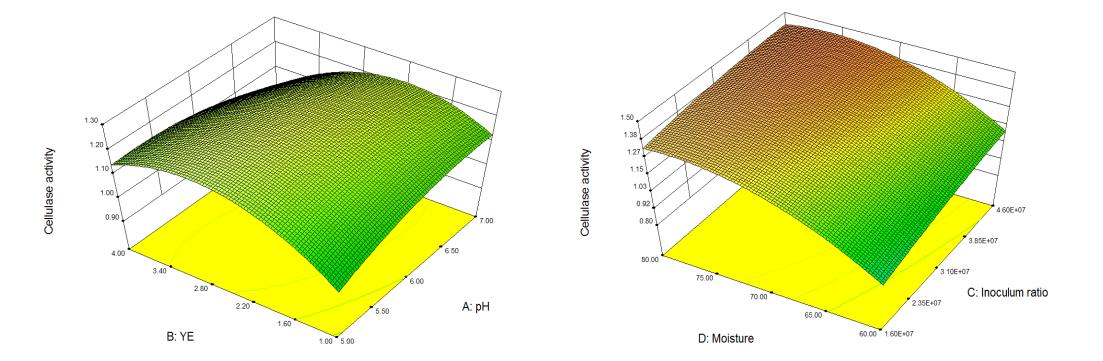
	Sum of		Mean	F	p-value	
Source	Squares	df	Square	Value	Prob > F	
Model	1.72	14	0.12	12.25	< 0.0001	significant
А-рН	4.778E-006	1	4.778E-006	4.755E-004	0.9829	
B-YE	4.431E-003	1	4.431E-003	0.44	0.5167	
C-Inoculum size	0.14	1	0.14	13.68	0.0021	
D-Moisture	0.60	1	0.60	59.30	< 0.0001	

#### Conclusions

- Model: significant
- Importance of factors:

Moisture > Inoculum size > > pH > Yeast extract

4. Optimization of fermentation condition by Response Surface Methodology



#### **Optimal condition**

Moisture 77-78%, inoculum size  $3.1 \times 10^7$  spores g<sup>-1</sup> textile,

pH 6-7, yeast extract 2.3 w/w%

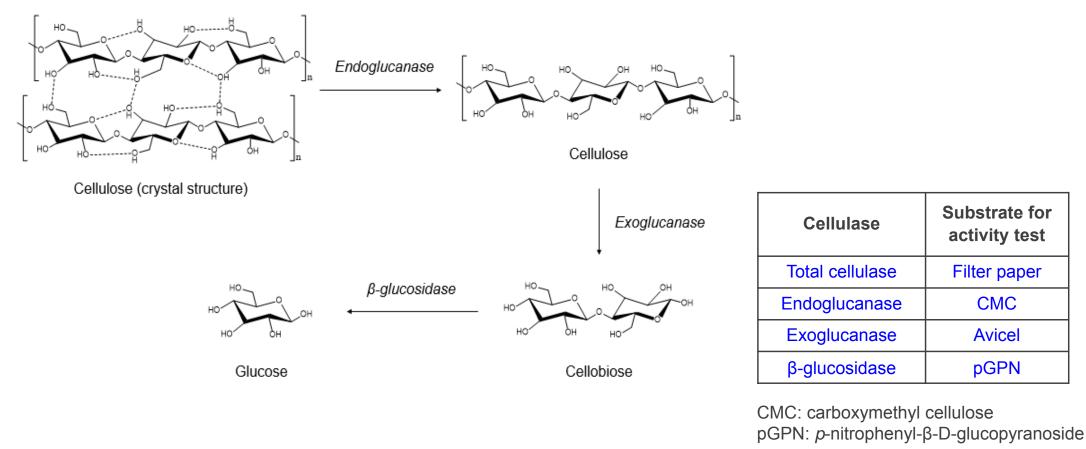
### **High point**

1.44 FPU g<sup>-1</sup> from cotton/PET 80/20

 $(17\% \text{ increase from } 1.24 \text{ FPU } \text{g}^{-1})$ 

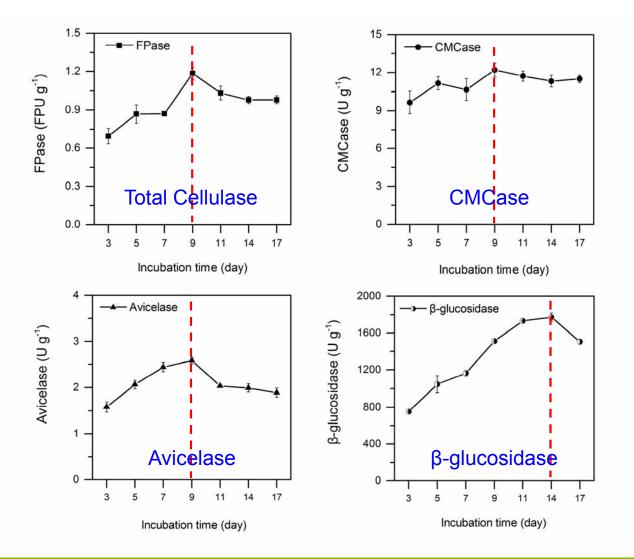
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5. Time courses of total cellulase activity and individual cellulase activities



Synergistic effect of a complete cellulase system

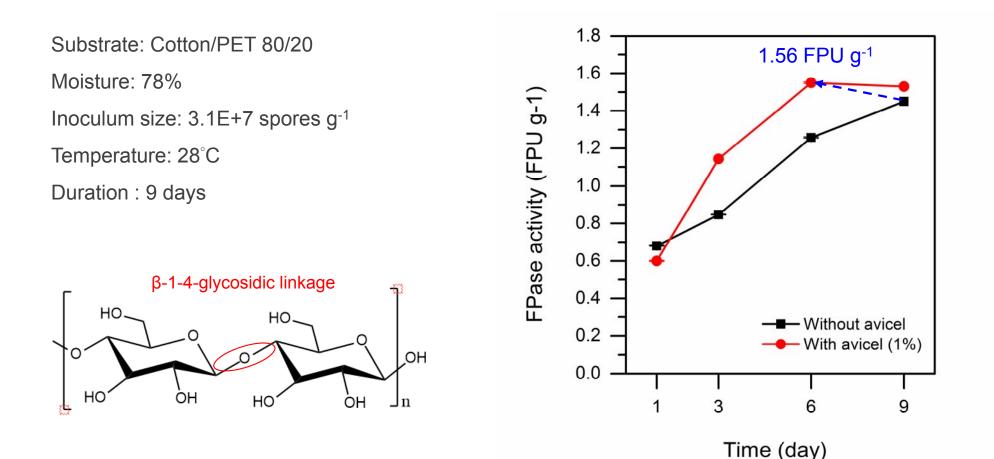
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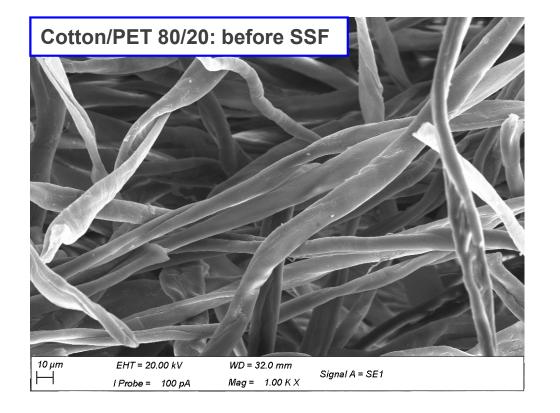
Effect of avicel (1 w/w%) on cellulase time profile

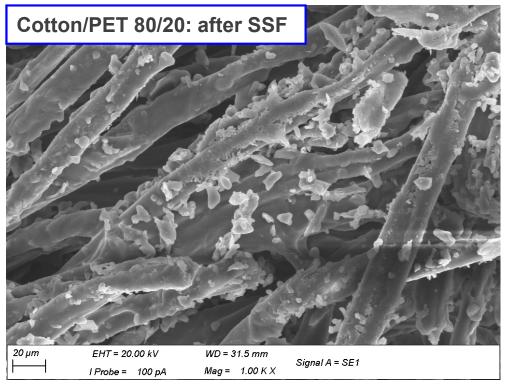


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6. Scanning Electron Microscope (SEM) detection

**Magnification of 1000x** 

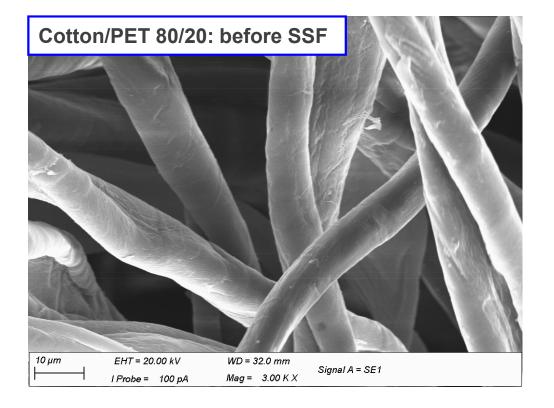


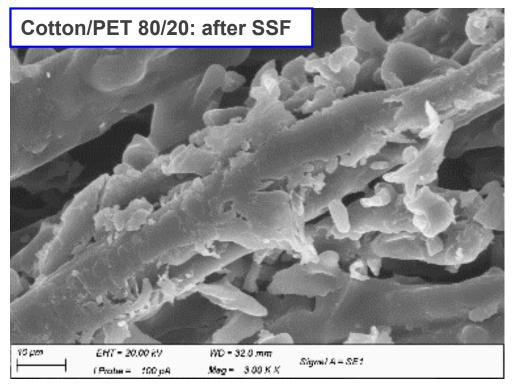


(ITP/109/15TP)

6. Scanning Electron Microscope (SEM) detection

**Magnification of 3000x** 



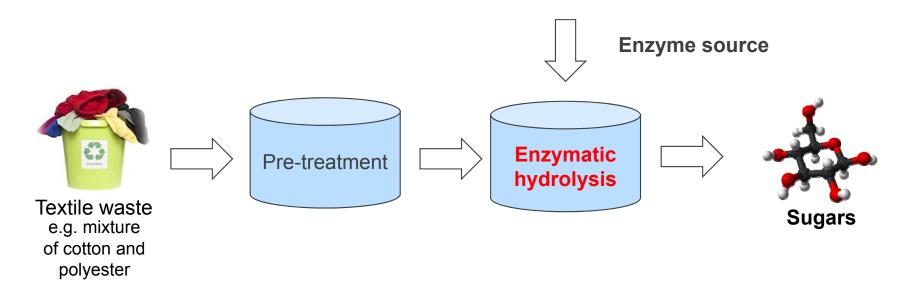


(ITP/109/15TP)

### **Research target**

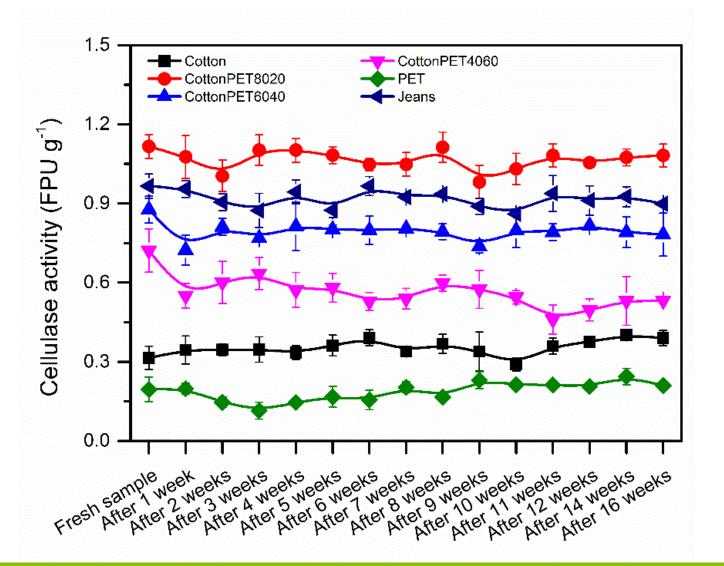
### Fungal cellulase production on textile waste

(Conducted by Dr. Carol Lin from CityU and Dr. Du Chenyu from the University of Huddersfield)



Materials	Dye	Pre-treatment	Source of enzymes
<ul> <li>100% Cotton</li> <li>100% Polyester</li> <li>Cotton/PET blend (80/20, 60/40, 40/60)</li> <li>Jean</li> </ul>	<ul><li>Reactive</li><li>Disperse</li><li>Indigo</li></ul>	<ul><li> Alkaline</li><li> Milling</li><li> Autoclave</li></ul>	<ul> <li>Fungal enzymes (solid state fermentation: SSF)</li> </ul>

#### 7. Retention of fungal cellulase extract (in -20°C freezer)



	Cellulase activity (FPU g <sup>-1</sup> )			
	Fresh sample	After 4 months		
Cotton	0.31	0.39		
Cotton/PET 80/20	1.12	1.08		
Cotton/PET 60/40	0.88	0.78		
Cotton/PET 40/60	0.72	0.53		
PET	0.20	0.21		
Jeans	0.97	0.90		

(ITP/109/15TP)

Cellulase activity maintained stably for 4 months

# Conclusions

- Cellulase was successfully produced on textile waste by solid state fermentation;
- The fungal strain *A. niger* CKB was selected (provided by Prof. Diannan Lu in Tsinghua University);
- <u>Moisture</u> and <u>inoculum size</u> are important factors;
- The highest cellulase activity was around 1.56 FPU g<sup>-1</sup>, obtained on textile of <u>cotton</u>
   <u>80% and PET 20%</u> within 6 days (80% moisture, 3.1E+7 spores g<sup>-1</sup> textile);
- Crystalline structure of textile substrate was partially disrupted by cellulase digestion;
- Retention of crude fungal cellulase activity by freezing storage for application in textile hydrolysis.

# Acknowledgements

- □ The Hong Kong Research Institute of Textiles and Apparel (HKRITA)
- □ **Sponsor:** H&M Hennes & Mauritz (Far East) Ltd.

### □ Supervisor:

• Dr. Carol Lin

### □ Collaborators:

- Dr. Chenyu Du (University of Huddersfield, United Kingdom)
- Dr. Shao-Yuan Leu (Hong Kong Polytechnic University)
- Dr. Hao Liu (South China University of Technology, China)
- Dr. Diannan Lu (Tsinghua University, China)

### Research team

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# Thank you for attention !

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### Literature review

	Treatment			Fermentation		Polyester	
Materials	Pre-treatment	Enzyme treatment	Glucose yield (%)	process	yield	separation Efficiency (%)	Ref.
white 40/60 polyester/cotton blend	12 wt% <mark>NaOH</mark> -20 ~ 0 °C  for 1h (5% solid load)	30 FPU cellulase 60 IU <mark>β-glucosidase</mark> per gram of cellulose	82	S. cerevisiae SSF	70% theoretical Ethanol	98	{Gholamzad, 2014 #1}
100% cotton linters; red T-shirt ; blue polyester/cotton	5 g/L Na2S2O4 and Na2CO3 solution 100 °C for 1 h	10 FPU/g <mark>Cellulase</mark> AP3 50 °C 250 rpm	90 80	Z. mobilis SSF	50 g/L Ethanol from 75g/L waste textile		{Kuo, 2014 #2}
(40/60) blended shirt	85% phosphoric acid 50 °C , 100 rpm for 2 h	48 h	60				
jeans	85% <mark>phosphoric acid</mark> 50 °C , 130 rpm for 24 h	7.5 FPU/g <mark>cellulase</mark> 50 °C 130 rpm 96 h	79.2			100	{Shen, 2013 #4}
100% cotton T-shirts	[AMIM]CI (ionic liquid) 110 °C for 90 min	66 U/g <mark>cellulase</mark> 50 ∘C, 80 rpm for 96 h	94				{Hong, 2012 #5}
orange 50/50 polyester/ cotton blend; blue 40/60 polyester/viscose blend	85% N- methylmorpholine-N- oxide 120 °C for 2h	20 FPU/g cellulase 30 IU/g <mark>β-glucosidase</mark> 48 h	92				{Jeihanipour , 2010 #6}
blue jeans textiles	18% (w/v) <mark>NaOH</mark> 23 °C for 3 h	20 FPU/g cellulase 30 IU β-glucosidase 45 °C for 48 h.	99	<i>S. cerevisia</i> e SSF	85–86%		{Jeihanipour , 2009 #7}

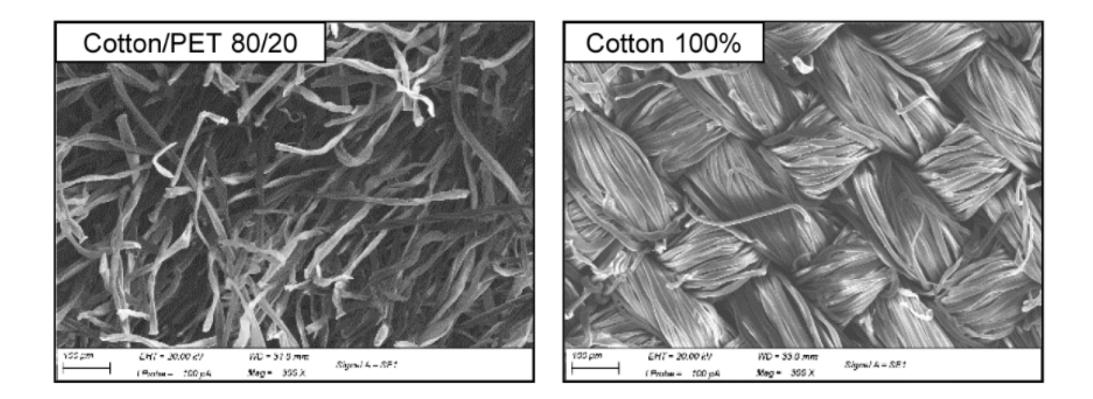
### **Analysis of Variance (F-test)**

 $F = \frac{Varience between treatment (MSTreatment)}{Varience within treatment (MSError)}$ 

$$MS_{Treatment} = \frac{\sum_{i} n_{i} (\overline{X}_{i} - \overline{X})^{2}}{K - 1}$$

$$MS_{Error} = \frac{\sum_{ij} n_i (X_{ij} - \overline{X}_i)^2}{N - K}$$

- N: number of all results obtained;
- K: number of treatment;
- n: number of result in a treatment;
- $\overline{X}$ : average value of all results;
- $\overline{X}_i$ : average value of results in a treatment



#### **Cotton/PET blended textile (furry surface structure)**

Larger surface area

Better aerobic condition

4. Optimization of fermentation condition by Response Surface Methodology

#### Suggested optimal solution

Moisture 78%, inoculum size  $4.6 \times 10^7$  spores g<sup>-1</sup> textile, pH 7.2-7.3, yeast extract 2.3 w/w%

#### Test

- 1) Inoculum size  $4.6 \times 10^7$  spores g<sup>-1</sup> textile:
- 2) Inoculum size  $3.1 \times 10^7$  spores g<sup>-1</sup> textile:
- 3) Inoculum size  $1.6 \times 10^7$  spores g<sup>-1</sup> textile:

Actual response 1.46 FPU g<sup>-1</sup> 1.44 FPU g<sup>-1</sup> 1.13 FPU g<sup>-1</sup>

**Predicted response** 

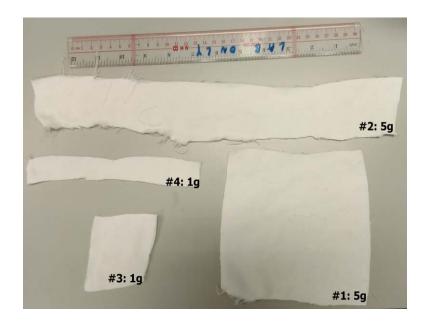
1.48 FPU  $g^{-1}$  (from cotton/PET 80/20)

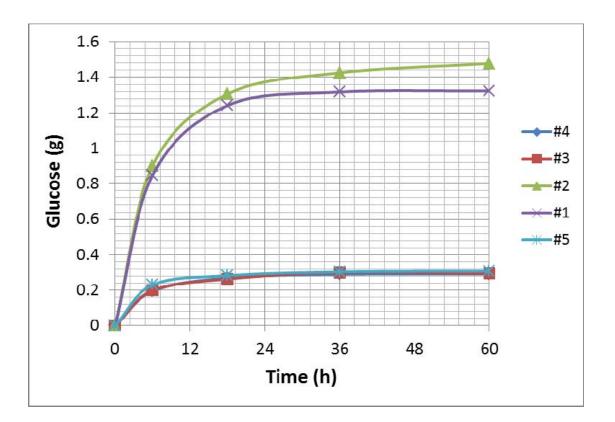
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### (ITP/109/15TP)

# Appendix

 Effect of fabric size on the enzymatic hydrolysis





No.	Shape	Initial weigh (g)	Final weigh (g)	Glucose (g)	Glucose recovery (g/kg)
1	Square $(14 \times 14 \text{ cm}^2)$	5.00	3.54	1.33	266
2	Rectangle ( $40 \times 5 \text{ cm}^2$ )	5.00	3.44	1.48	296
3	Square $(6.3 \times 6.3 \text{ cm}^2)$	1.00	0.66	0.29	290
4	Rectangle ( $18 \times 2.2 \text{ cm}^2$ )	1.00	0.70	0.29	290
5	Pieces $(0.3 \times 0.3 \text{ cm}^2)$	1.00	0.67	0.31	310

### (ITP/109/15TP)

# Appendix

### **Cellulase activity measurement**

#### **Enzyme extraction**

(N. Pensupa, 2013)

- 1) Dissolved in 5 mM citric acid buffer
- 2) Blending and centrifuge
- 3) Collect the enzyme solution





#### Cellulase activity detection

(Filter paper activity, B. Adney, 1996)

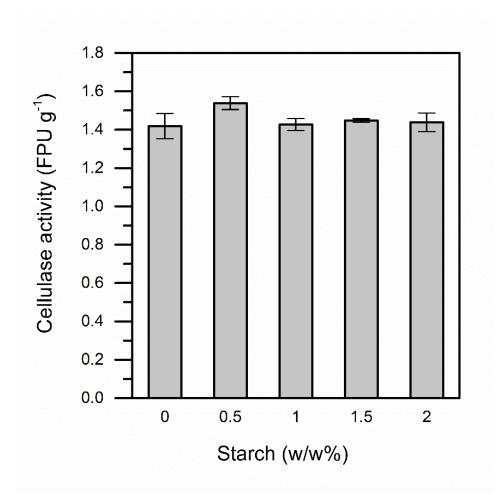
1) 0.5 ml enzyme solution + 1 ml citric acid buffer + 50 mg filter paper roll

- 2) Water bath 50 °C for 1 h
- 3) + 3 ml DNS solution and boiling for 5 minutes
- 4) UV detection at 540 nm

### (ITP/109/15TP)

#### Effect of starch on SSF

Moisture: 80%, Temperature: 28 °C, Duration : 9 days, Substrate: Cotton/PET 80/20



Starch as carbon source to support cell growth

Starch	Cellulase activity	REF
0% to 0.5%	1.42 to 1.53 FPU (increase 7.7%)	This study
0% to 0.75%	0.76 to 0.87 FPU (increase 14%)	Liang et al, 2012



# Conclusion: insignificant

Effect of different cellulase producing inducers

Moisture: 80%, Temperature: 28 °C, Duration : 9 days, Substrate: Cotton/PET 80/20

Loading	Cellulase activity (FPU g <sup>-1</sup> )					
ratio (w/w %)	Sucrose	Lactose	CMC	Avicel	Filter paper scrap	
0	1.54	1.49	1.52	1.53	1.50	
0.5	1.52	1.48	1.14	1.52	1.37	
1	1.39	1.37	0.81	1.55	1.18 <b>1.3%</b>	
1.5	1.42	1.06	0.98	1.47	1.07	
2	1.43	1.17	1.07	1.40	1.28	
5	1.27	1.16	1.05	1.42	1.24	

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