Biodegradation of phenolic effluent of producer gas plant using Scenedesmus sp.

IWWATV-2015

Dr. A.K. Bakthavatsalam, Professor of Energy & Head of T&P, NIT-Tiruchirappalli, Tamilnadu, India. 5/29/2015



Need for the work

Acute exposure of phenol can result in myocardial depression, muscle weakness gastrointestinal disturbance, tremors, skin whitening, corneal whitening and finally blindness and even damage central nervous system.(Public Health Statement)

Replayer of the terminal of the terminal termina

Realt would be useful to the industries viz. coal chemical plants, refineries, petrochemical industries, fibre glass units, explosive manufacture, polymerization process, pharmaceuticals, plastic, paints, textile units making use of organic dyes, biocides, photographic chemicals etc.

Outline

Real Introduction

- Available methods of treatment from literature
- Real Background of study
- Reprocess flow chart
- Characterization of raw effluent

Selection and cultivation of targeted algal species
Experimentation
Analysis of Phenol
Result and Discussion
Conclusion
Acknowledgements
References

Introduction

- It contains a variety of combustible gases including H_2 , CO, CH_4 and volatile hydrocarbons together with small quantities of non-calorific gases such as CO_2 and N_2 .
- Reproduct from the producer gas plant includes coal tar and ammonia.

In producer gas plant,

- 200m³/Day of water is used for coal gasification and gas cleaning system.
- ☑ 150m³ / Day of water returns as phenolic effluent.
- ☑ This waste water contains phenolic compounds, tar and oil. Around 0.05m³ of tar is collected per day using tar oil separator.

Introduction- contd....

Area of Research :

Phycoremediation of Phenolic Effluent of Producer Gas Plant .

Source of sample:



Producer Gas Plant, Bharath Heavy Electrical (BHEL),Trichy, Tamilnadu, India.

Available methods of treatment from literature

Identified methods of treatment from literature:

- Chemical Degradation
- Biodegradation
- Degradation using UV
- Enzymatic Degradation
- Physical techniques

Algae used for the treatment of phenol and its types

Types of phenol	Name of microlagae	Reference		
Nonylphenol	C. caspia, Chlorella vulgaris, Selenastrum capricornutum, Chlorella sp.	 Yu, Liu1, Dai, Xiaokang., Wei, Jie., 2013. Toxicity of the xenoestrogen nonylphenol and its biodegradation by the alga <i>Cyclotella caspia</i>. Journal of Environmental Sciences, 25(8) 1662–1671. Gao, Q.T., Wong, Y.S., Tam, N.F.Y., 2011, Removal and biodegradation of nonylphenol by immobilized <i>Chlorella vulgaris</i>, Bioresource Technology 102:10230–10238. Gao, Q.T., Tam, N.F.Y., 2011, Growth, photosynthesis and antioxidant responses of two microalgal species, <i>Chlorella vulgaris</i> and <i>Selenastrum capricornutum</i>, to nonylphenol stress, Chemosphere 82:346–354. Gao,Q.T., Wong, Y.S., Tam, N.F.Y., 2011. Removal and biodegradation of nonylphenol by different <i>Chlorella</i> species; Marine Pollution Bulletin. 63 :445–451. 		
Nonylphenol, Octylphenol	Scenedesmus obliqus	Zhou,G.J., Peng, Fu-Qiang., Yang, Bin., Ying, Guang-Guo., 2013. Cellular responses and bioremoval of nonylphenol and octylphenol in the freshwater green microalga <i>Scenedesmus obliquus</i> , Ecotoxicology and Environmental Safety 87:10–16		
Chlorophenol	Chlorella VT-1	Olivier, S., Scragg, A.H., Morrison, J., 2003. The effect of chlorophenols on the growth of <i>Chlorella</i> VT-1, Enzyme and Microbial Technology 32 :837–842.		
Phenol	Phormidium valderianum, Ochromonas danica, Chlorella VT-1	Shashirekha, S., Uma,L., Subramanian, G., 1997. Phenol degradation by the marine cyanobacterium <i>Phormidium valderianum</i> BDU 30501, J. Indust. Microbiol. and Biotech. 19(2): 130-133. Scragg, A.H., 2006. The effect of phenol on the growth of <i>Chlorella vulgaris</i> and <i>Chlorella</i> VT-1, Enzyme and Microbial Technology 39, 796–799.		

Process flow chart of gas cooling system of producer gas plant



Effluent Treatment Plant

Characterization of raw effluent

601

Parameters and the standard testing procedures

Parameter	Test procedure	Range limit		
Colour	IS: 3025 (Part-4) :1983/ APHA 22nd -2120B-2012	1 to 50 Hazen		
Oil & Grease	IS: 3025 (Part-39): 1991/ APHA 22nd ed5520-2012	5 to 1000 mg/l		
pH	IS: 3025 (Part-11) :1983/ APHA 22nd ed2500-2012	1 to 14		
TDS	IS: 3025 (Part-16) :1984/APHA 22nd ed2541-2012	1 to 20000 mg/1		
TSS	IS: 3025 (Part-17) :1984/ APHA 22nd ed2540-2012	1 to 1000 mg/1		
Phenolic Compounds (as C ₆ H ₅ OH)	IS: 3025 (Part-43) : 1992/RA2003	0.02 to 150 mg/1		
Biochemical Oxygen Demand (BOD)	IS: 3025 (Part-44) : 1993/ APHA 22nd ed5210B-2012	1 to 10000 mg/1		
ChemicalOxygenDemand (COD)	IS: 3025 (Part-58) : 2006/ APHA 22nd ed5220-2012	1 to 20000 mg/l		

5/29/2015

Characterization of raw effluent- contd......

Initial concentration of conventional parameters and Total phenol concentration of raw effluent

(Total Phenol concentration was determined by IS 3025 part no 43)

Parameter	Units	Mean ± SD
Total suspended solids	mg/l	244.75 ±48.7
Total dissolved solids	mg/l	827.4 ±92.1
Biological oxygen demand	mg/l	62.5 ±12.3
Chemical oxygen demand	mg/l	2116 ±119.9
Oil & grease	mg/l	97.25 ±13.5
Phenolic concentration	mg/l	1024±21.8
рН	8-9.2	
Color	Dark Brown	

Analysis of types of phenolic compounds

Method:

- The phenolic compounds was extracted from the effluent using Ethyl acetate as a solvent.
- Restracted samples were analyzed in GC using the following operating conditions.

Standards selected based on EPA:

- 1. Phenol
- 2. 2-Chlorophenol
- 3. 2-Methylphenol
- 4. 2-Nitrophenol
- 5. 2,4-Dimethylphenol
- 6. 2,4-Dichlorophenol
- 7. 4-Chloro-3-methylphenol
- 8. 2,4,6-Trichlorophenol
- 9. 2,4-Dinitrophenol
- 10. 4-Nitrophenol
- 11. Pentachlorophenol

Reference:

Handbook of Water Analysis, Second Edition, edited by Leo M.L. Nollet, Leen S. P. De Gelder.

5/29/2015

Analysis of types of phenol present in the raw effluent

(using Gas Chromatography)



Selection and cultivation of targeted algal species

Source of sample:

Procured from the Bioenergy lab, Department of Energy and environment, National Institute of Technology, Trichy, India.

Cultivation of *Scenedesmus* **sp.** :

The *Scenedesmus* sp. was cultivated in optimized culture medium comprising potassium bicarbonate and urea in the ratio of 2:1.

Culture condition- Atmospheric

Temperature range - 30 - 35° C.

рН-7-8.

The growth of microalgae was monitored by observing optical density at 600 nm and for the identification of contamination, the algae was monitored daily in fluorescent inverted microscope (Nikon DS-Fi2).

Experimentation

- A The 15 day old culture was centrifuged and four different concentrations (1g, 2g, 3g and 4g per liter of raw effluent) of wet biomass with nutrients (Potassium bicarbonate: Urea in the ratio of 2:1) (Batch I) and without nutrients (Batch II) was inoculated into raw effluent
- Samples and controls were monitored for 7 days at ambient temperature and environmental conditions.
- A The algal growth was monitored by analyzing optical density at 600 nm using UV-VIS spectrophotometer. Fluorescent Inverted Microscope was used to monitor physical changes in the algae.
- ↔ The phenolic concentration was analyzed using IS 3025 Part no: 43 methods.





Result and Discussion



Effect of phenol on the growth of Scenedesmus sp.



Effect of phenol on the growth of *Scenedesmus* sp.



5/29/2015

Reduction of Total phenol



Degradation of Phenol(with nutrients)



ppm- mg of algal wet biomass / liter of effluent



Degradation of Phenol (with nutrients)



Degradation of Phenol (without nutrients)



ppm- mg of algal wet biomass / liter of effluent



Reduction of carbon content

Effect of *Scenedesmus* sp. on the carbon content (Organic Carbon (OC), Inorganic Carbon (IC), Total Carbon (TC)) of batch-I



Effect of *Scenedesmus* sp. on the carbon content (Organic Carbon (TOC), Inorganic Carbon (IC), Total Carbon (TC)) of batch-II



Conclusion

- R Phycoremediation of waste water treatment integrated with conventional methods present great opportunities to the water and wastewater treatment technologies.

- Removal of contaminants present in the effluent other than phenol could also be noticed when the *Scenedesmus* sp. was externally supplied with nutrients.

Acknowledgements

Authors would like to thank the **National Institute of Technology, Trichy** and the World Bank funded **Technical Education Quality Improvement Program (TEQIP)**.

References

- Klekner, V., Kosaric, N., 1992. Degradation of phenols by algae. Environmental Technology.13:493-501.
- Semple, K.T., Cain, R.B., 1996. Biodegradation of Phenols by the Alga *Ochromonas Danica*, Applied and Environmental Microbiology, 1265–1273.
- Semple , K.T ., Cain, R.B., Schmidt, S., 1999. Biodegradation of aromatic compounds by microalgae, FEMS Microbiology Letters.170: 291-300.
- Pinto, G., Pollio, A., Previtera, L., Temussi, F., 2002. Biodegradation of phenol, Biotechnology Letters. 24:2047-2051.
- Hirooka, H., Akiyama, Y., Tsuji, N., Nakamura, T., Nagase, H, Hirata, K., Miyamoto, K., 2003. Removal of Hazardous Phenols by Microalgae under Photoautotrophic Conditions, Journal of bioscience and bioengineering. 95(2):200-203.
- C Lima, Sofia A.C., Castro, Paula M.L. and Morais, R., 2003, Biodegradation of *p*-nitrophenol by microalgae. Journal of applied phycology. 15(3):137-142.
- Lima, S.A.C., Raposo, M.F.J., Castro, P.M.L., Morais, R.M., 2004. Biodegradation of p-chlorophenol by microalgae consortium. 38(1): 97-102.
- Kumar, A., Kumar, S., and Kumar, S., 2005. Biodegradation kinetics of phenol and catechol using *Pseudomonas putida* MTCC 1194. Biochemical Engineering Journal. 22: 151–159.
- Melo, P.S., Fabrin-Neto, J.B., De Moraes, S.G., Assalin, M.R., Duran, N., Haun. M., 2006. Comparative toxicity of effluents processed by different treatments in V79 fibroblasts and the Algae *Selenastrum capricornutum*. Chemosphere. 62:1207–121.

5/29/2015

References-contd....

- Scragg, A.H., 2006. The effect of phenol on the growth of *Chlorella vulgaris* and *Chlorella* VT-1, Enzyme and Microbial Technology. 39:796–799.
- Michałowicz, J., Duda, W., 2007. Phenols transformations in the environment and living organisms, Current Topics in Biophysics. 30(suppl. A): 24-36.
- Retroutsos. D., Katapodis, P., Christakopoulos, P., Kekos, D., 2007, Removal of p-chlorophenol by the marine microalga *Tetraselmis marina*, Journal of Applied Phycology, 19:485–490.
- Edalatmanesh, M., Mehrvar, M., Dhib, R. 2008. Optimization of phenol degradation in a combined photochemicalbiological wastewater treatment system. Chemical Engineering Research and Design. 86:1243–1252.
- № Lika, K., Papadakis, I. A., 2009. Modeling the biodegradation of phenolic compounds by microalgae, Journal of Sea Research, 62:135–146.
- Christenson, L., Sims, R., 2011. Production and harvesting of microalgae for wastewater treatment, biofuels, and bioproducts. Biotechnology Advances. 29:686–702.
- Ganesamoorthy, S., Jerome, P., Shanmugasundaram, K., and Karvembu, R., 2014. Highly efficient homogeneous and heterogenized ruthenium catalysts for transfer hydrogenation of carbonyl compounds, RSC Advances. 4:27955-27962.
- Komolafe, O., Orta, S.B.V., Monje-Ramirez, I., Noguez, I.Y., Harvey, A.P., Ledesma, M.T.O., 2014. Biodiesel production from indigenous microalgae grown in wastewater, Bioresource Technology.154: 297–304.
- Nollet, L.M.L., De Gelder, L.S. P., Handbook of Water Analysis, Second Edition, ISBN-13: 978-1439889640.

5/29/2015



 $(\gamma$

